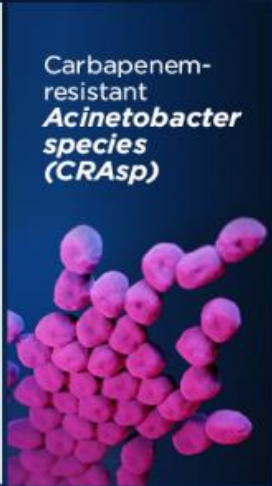


MDRO Reporting and Investigation in Michigan

6 of the 18 most alarming **antibiotic resistance threats** cost the U.S. more than **\$4.6 billion annually**



Vancomycin-resistant
Enterococcus
(VRE)



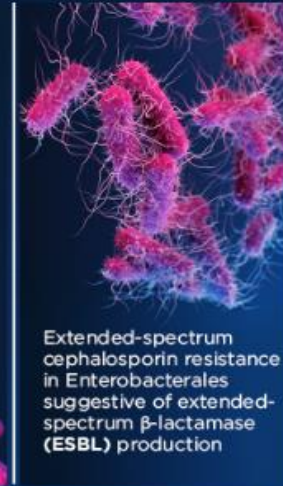
Carbapenem-resistant
Acinetobacter
species
(CRAsp)



Methicillin-resistant
Staphylococcus
aureus (MRSA)



Carbapenem-resistant
Enterobacterales
(CRE)



Extended-spectrum
cephalosporin resistance
in *Enterobacterales*
suggestive of extended-
spectrum β -lactamase
(ESBL) production



Multidrug-resistant (MDR)
Pseudomonas
aeruginosa

www.cdc.gov/DrugResistance



U.S. Department of
Health and Human Services
Centers for Disease
Control and Prevention

Niki Mach, MPH, CPH, MT(ASCP)

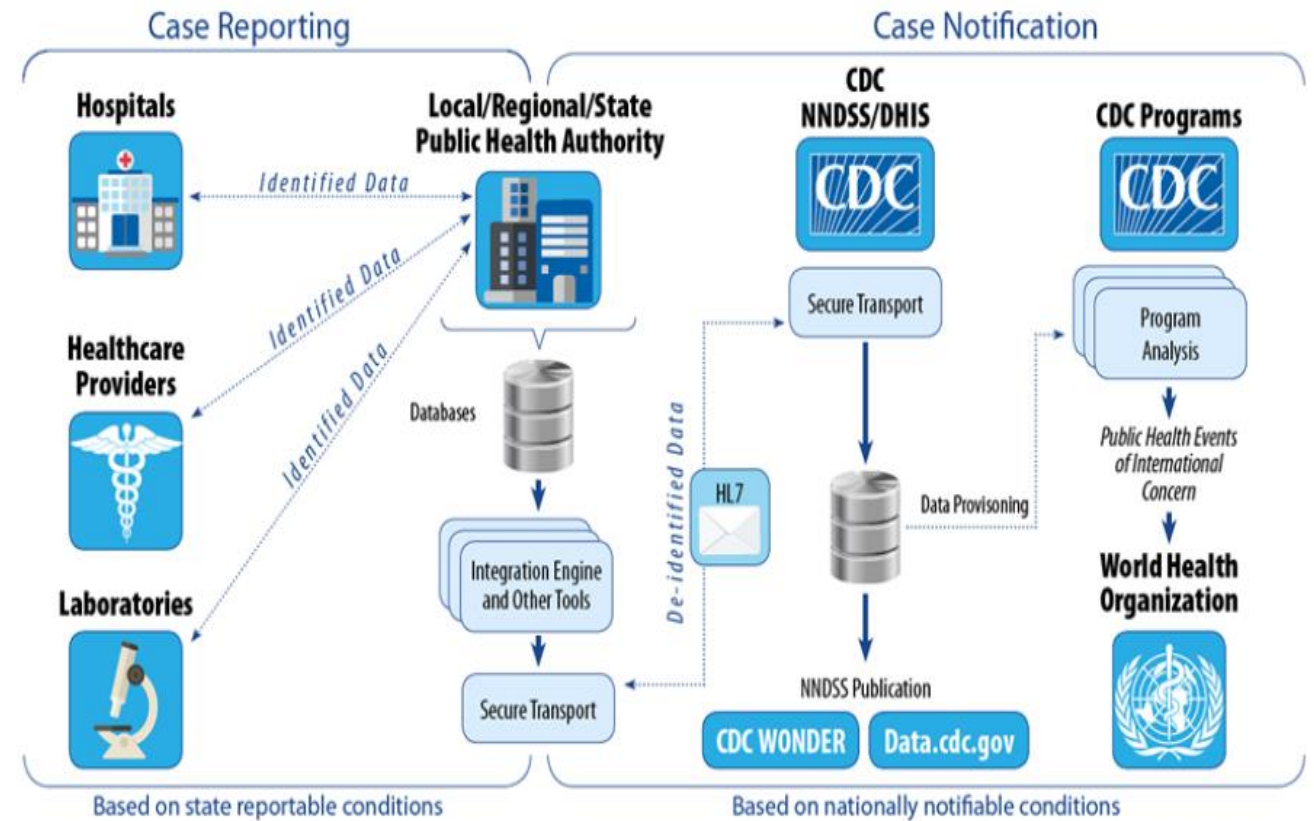
Margaret Sturgis, MSA, BSN, RN

Surveillance for Healthcare-Associated and Resistant Pathogens (SHARP) Unit [Staff](#)

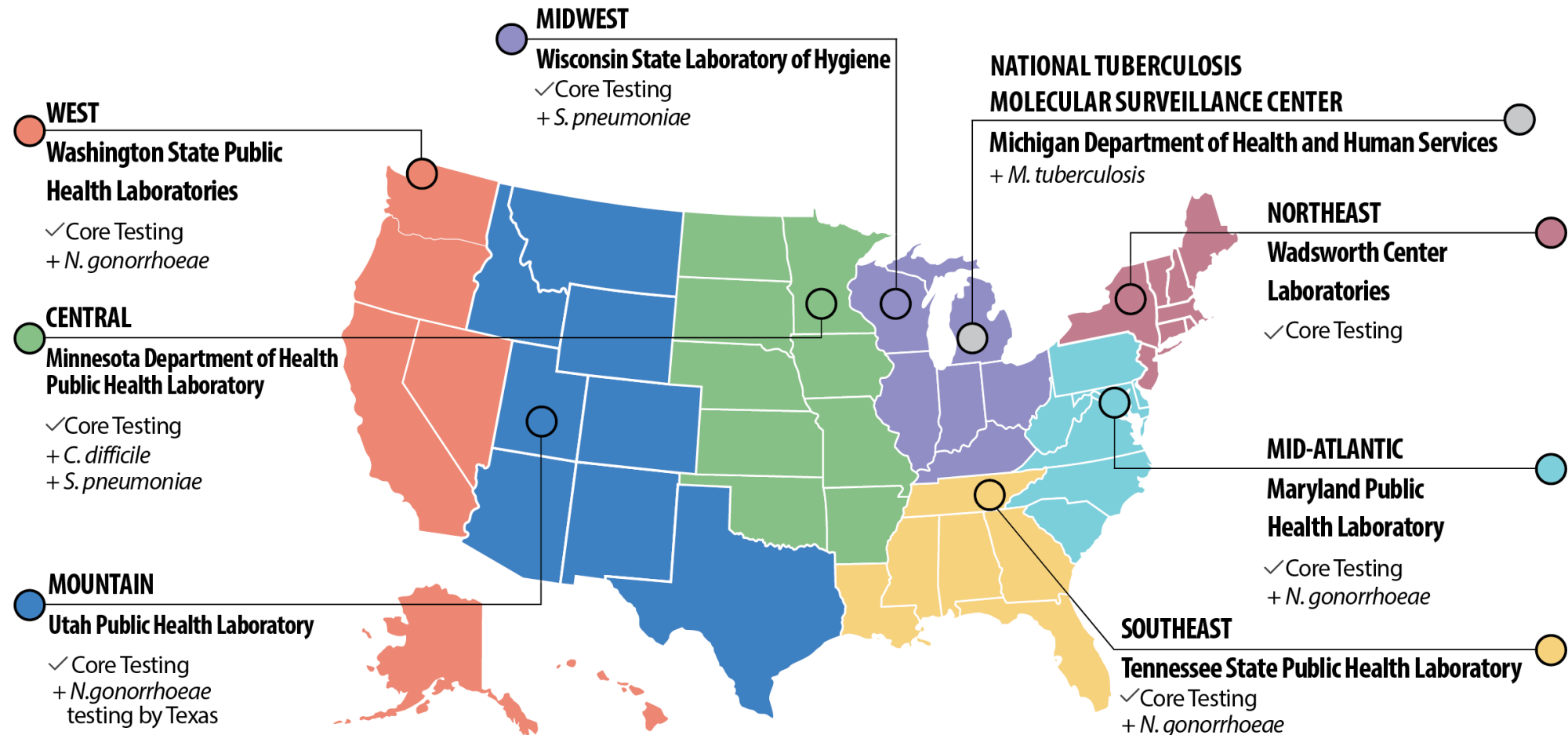
Michigan Department of Health and Human Services

Reportable Diseases in Michigan

- Michigan Disease Surveillance System (MDSS) is the state database for collecting surveillance data.
 - Web-based communicable disease reporting system
 - Cases can be reported by:
 - Electronic laboratory report (ELR)
 - Manual case entry
- Required case reporting to MDSS by healthcare providers and laboratories
- [Surveillance case definition](#) endorsed by CSTE/CDC, nationally notifiable



Antibiotic Resistance Laboratory Network



Antimicrobial Resistant Reportable Diseases

- *Candida auris* (Candidiasis)
- Carbapenem-Producing, Carbapenem-Resistant *Enterobacterales* (CP-CRE). Reportable in MI starting January 2018
 - **CP-CRE Case Surveillance**
 - Required case reporting to MDSS by healthcare providers and laboratories
 - Carbapenemase producing – carbapenem resistant *Enterobacterales (All Genera)*
 - **CP-CRE Isolate Surveillance**
 - Required isolate submission to BOL by laboratories
 - Carbapenemase-producing – carbapenem resistant *Enterobacterales (All Genera)*
- *Staphylococcus aureus*, Vancomycin Intermediate/Resistant (VISA/VRSA)
- Unusual occurrence, outbreak, or epidemic

NEW for
2022

2022 REPORTABLE DISEASES IN MICHIGAN – BY PATHOGEN A Guide for Physicians, Health Care Providers and Laboratories

Report the following conditions to the Michigan Disease Surveillance System (MDSS) or local health department (see reverse) within 24 hours if the agent is identified by clinical or laboratory diagnosis. See footnotes for exceptions.

Report the unusual occurrence, outbreak or epidemic of any disease or condition, including healthcare-associated infections.

Acute flaccid myelitis (1) Anaplasma phagocytophilum (Anaplasmosis) Arboviral encephalitis, neuro- and non-neuroinvasive: Chikungunya, Eastern Equine, Jamestown Canyon, La Crosse, Powassan, St. Louis, West Nile, Western Equine, Zika (6) Babesia microti (Babesiosis) Bacillus anthracis and B. cereus serovar anthracis (Anthrax) (4) Blastomycosis dermatitis (Blastomycosis) Bordetella pertussis (Pertussis) Borrelia burgdorferi (Lyme Disease) Brucella species (Brucellosis) (4) Burkholderia mallei (Glanders) (4) Burkholderia pseudomallei (Melioidosis) (4) Campylobacter species (Campylobacteriosis) Candida auris (Candidiasis) (4) Carbapenemase Producing – Carbapenem Resistant Enterobacterales (CP-CRE): all genera (4) Chlamydia trachomatis (Trachoma, genital infections, LGV) (3, 6) Chlamydia pneumoniae (Psittacosis) Clostridium botulinum (Botulism) (4) Clostridium tetani (Tetanus) Coccidioides immitis (Coccidioidomycosis) Coronaviruses, Novel; including deaths and SARS-CoV-2 variant identification (SARS, MERS-CoV, SARS-CoV-2) (5) Corynebacterium diphtheriae (Diphtheria) (5) Coxiella burnetii (Q Fever) (4) Cronobacter sakazakii (4, blood or CSF only, from infants < 1 year of age) Cryptosporidium species (Cryptosporidiosis) Cyclospora species (Cyclosporiasis) (5) Dengue virus (Dengue Fever) Ehrlichia species (Ehrlichiosis) Encephalitis, viral or unspecified Escherichia coli, O157:H7 and all other Shiga toxin positive serotypes (including HUS) (5) Francisella tularensis (Tularemia) (4) Giardia species (Giardiasis) Guillain-Barre Syndrome (1) Haemophilus ducreyi (Chancroid) Haemophilus influenzae, sterile sites (5, submit isolates for serotyping for patients <15 years of age) Hantavirus Hemorrhagic Fever Viruses (4) Hepatitis A virus (Anti-HAV IgM, HAV genotype) Hepatitis B virus (HBsAg, HBeAg, anti-HBc IgM, HBV NAAT, HBV genotype; report all HBsAg and anti-HBs (positive, negative, indeterminate) for children ≤ 5 years of age) (6) Hepatitis C virus (all HCV test results including positive and negative antibody, RNA, and genotype tests) (6) Histoplasma capsulatum (Histoplasmosis) HIV (tests including: reactive immunoassays (e.g., Ab/Ag, TD1/TD2, WB, EIA, IA), detection tests (e.g., VL, NAAT, p24, genotypes), CD4 counts/percents; and all tests related to perinatal exposures) (2, 6) Influenza virus (weekly aggregate counts) Pediatric influenza mortality, report individual cases (5) Novel influenza viruses, report individual cases (5, 6) Kawasaki Disease (1) Legionella species (Legionellosis) (5)	Leptospirosis (Leptospirosis) Listeria monocytogenes (Listeriosis) (5, 6) Measles virus (Measles/Rubeola) (6) Meningitis: bacterial, viral, fungal, parasitic, and amebic Multisystem Inflammatory Syndrome in Children (MIS-C) and in Adults (MIS-A) Mumps virus Mycobacterium leprae (Leprosy or Hansen's Disease) Mycobacterium tuberculosis complex (Tuberculosis); report preliminary and final rapid test and culture results (4) Neisseria gonorrhoeae (Gonorrhea) (3, 6) (4, submit isolates from sterile sites only) Neisseria meningitidis, sterile sites (Meningococcal Disease) (5) Orthopox viruses, including: Smallpox, Monkeypox (4) Plasmodium species (Malaria) Poliovirus (Polio) Prion disease, including CJD Rabies virus (4) Rabies: potential exposure and post exposure prophylaxis (PEP) Rickettsia species (Spotted Fever) Rubella virus (6) Salmonella species (Salmonellosis) (5) Salmonella Paratyphi (Paratyphoid Fever): serotypes Paratyphi A, Paratyphi B (tartrate negative), and Paratyphi C (5) Salmonella typhi (Typhoid Fever) (5) Shigella species (Shigellosis) (5) Staphylococcus aureus Toxic Shock Syndrome (1) Staphylococcus aureus, vancomycin intermediate/ resistant (VISA) (5)/VRSA (4) Streptococcus pneumoniae, sterile sites Streptococcus pyogenes, group A, sterile sites, including Streptococcal Toxic Shock Syndrome (STSS) Treponema pallidum (Syphilis) (6) Trichinella spiralis (Trichinellosis) Varicella-zoster virus (Chickenpox) (6) Vibrio cholera (Cholera) (4) Vibrio species (Vibriosis: non-cholera species) (5) Yellow fever virus Yersinia enterocolitica (Yersiniosis) (5) Yersinia pestis (Plague) (4)
--	--

LEGEND

- (1) Reporting within 3 days is required.
 - (2) Report HIV labs electronically/by arrangement & case reports by MDHHS Form 1355. Report HIV genome sequence data only as Sanger sequences, or as consensus sequences for next generation sequencing.
 - (3) Sexually transmitted infection for which expedited partner therapy is authorized. See www.michigan.gov/hivstl for details.
 - (4) A laboratory shall immediately submit suspect or confirmed isolates, subcultures, or specimens from the patient being tested to the MDHHS Lansing laboratory.
 - (5) Isolate requested. Enteric: If an isolate is not available from non-culture based testing, the positive broth and/or stool in transport medium must be submitted to the MDHHS Lansing laboratory.
 - (6) Report pregnancy status, if available.
- Blue Bold Text = Category A Bioterrorism or Select Agent must be notified immediately to the MDHHS Laboratory (517-335-8063)

This reporting is expressly allowed under HIPAA and required by Michigan Public Act 368 of 1978, 333.5111
 MDHHS maintains, reviews, and revises this list at least annually, for the most recent version please refer to: www.michigan.gov/cdinfo
 Michigan Department of Health and Human Services • Bureau of Laboratories • Bureau of Infectious Disease Prevention

REV. 7/2022

<https://www.michigan.gov/cdinfo>

2022 Brick Book and CD Lists

Candida auris Reporting Requirements

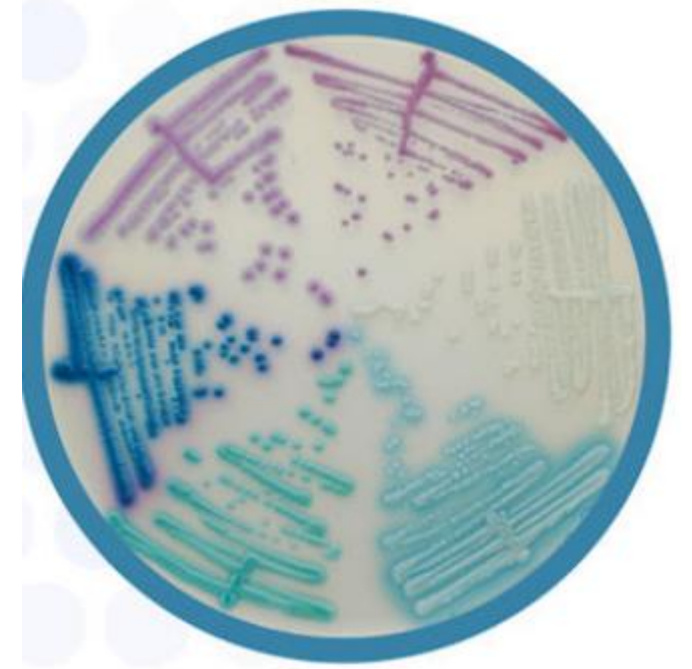
- Report any laboratory finding that meets either of the following criteria:
 - Detection of *C. auris* in a specimen using either culture or a culture-independent diagnostic test (CIDT) (e.g., Polymerase Chain Reaction [PCR])
 - Detection of an organism that commonly represents a *C. auris* misidentification in a specimen by culture (i.e., *Candida haemulonii*)
- Laboratories **shall immediately submit confirmed or suspect *C. auris*** isolates, subcultures, or specimens from the patient being tested to the MDHHS Lansing laboratory



Candida auris Case Report

Confirmatory laboratory evidence:

- Detection of *C. auris* from any body site using either culture or a culture independent diagnostic test (CIDT) (e.g., Polymerase Chain Reaction [PCR]).



Lab Results					
Report Date (mm/dd/yyyy)	Test Name	Reported Test Name/Test Result		Specimen	Collection Date (mm/dd/yyyy)
03/25/2022	Fungal Identification	Fungus identified/null	Candida auris///	Ear sample	05/20/2021
03/25/2022	Fungus identified	Fungus identified/Fungal Cultural Human	Candida auris///		05/20/2021
06/02/2021	Culture and Gram Stain Ear	BACTERIA IDENTIFIED: PRID:PT:EAR:NOM:AER OBIC CULTURE/Culture and Gram Stain Ear	Candida auris///	Ear sample	05/20/2021
05/25/2021	Culture and Gram Stain Ear	BACTERIA IDENTIFIED: PRID:PT:EAR:NOM:AER OBIC CULTURE/Culture and Gram Stain Ear	Candida auris///	Ear sample	05/20/2021
05/20/2021	Bacteria Identification [Presence] in Isolate by Culture	Bacteria Identification [Presence] in Isolate by Culture	//with normal skin flora CANDIDA AURIS Quanti ty of Organism: MODER ATE/		05/24/2021





2022 CP-CRE Case Reporting to MDSS



Physicians and laboratories **must report cases** of CP-CRE:

- ✓ Healthcare record contains a **diagnosis** of **Carbapenemase-producing Carbapenem-resistant Enterobacterales (CP-CRE)**, with KPC, NDM, OXA-48, IMP, VIM or a novel carbapenemase
- ✓ **Any Enterobacterales isolate** demonstrating carbapenemase production by a **phenotypic** test (e.g., Carba NP, CIM, mCIM)
- ✓ **Any Enterobacterales isolate** with a known carbapenemase resistance mechanism by a recognized **molecular test** (e.g., PCR, Expert Carba-R) for *Klebsiella pneumoniae* carbapenemase (KPC), New Delhi metallo- β -lactamase (NDM), Verona integron encoded metallo- β -lactamase (VIM), Imipenemase metallo- β -lactamase (IMP), Oxacillinase-48 (OXA-48)
- ✓ If testing for carbapenemase production (phenotypic) or carbapenemase resistance mechanism (molecular test) was not conducted or reported, **any Enterobacterales** isolate with a **minimum inhibitory concentration** of ≥ 4 mcg/ml for meropenem, imipenem, or doripenem, or ≥ 2 mcg/ml for ertapenem by antimicrobial susceptibility testing
 - ✓ *Morganella*, *Proteus*, *Providencia* spp. may have intrinsic resistance to imipenem. Only those isolates that are resistant to 1 or more carbapenems other than imipenem should be reported.

NEW for
2022

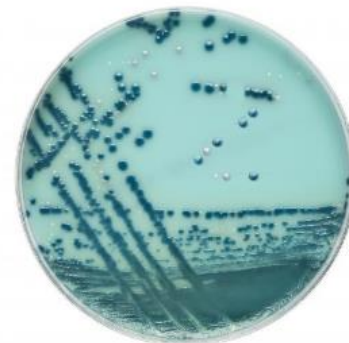
2022 CP-CRE Isolate Submission to BOL

Laboratories **must submit isolates** of CP-CRE:

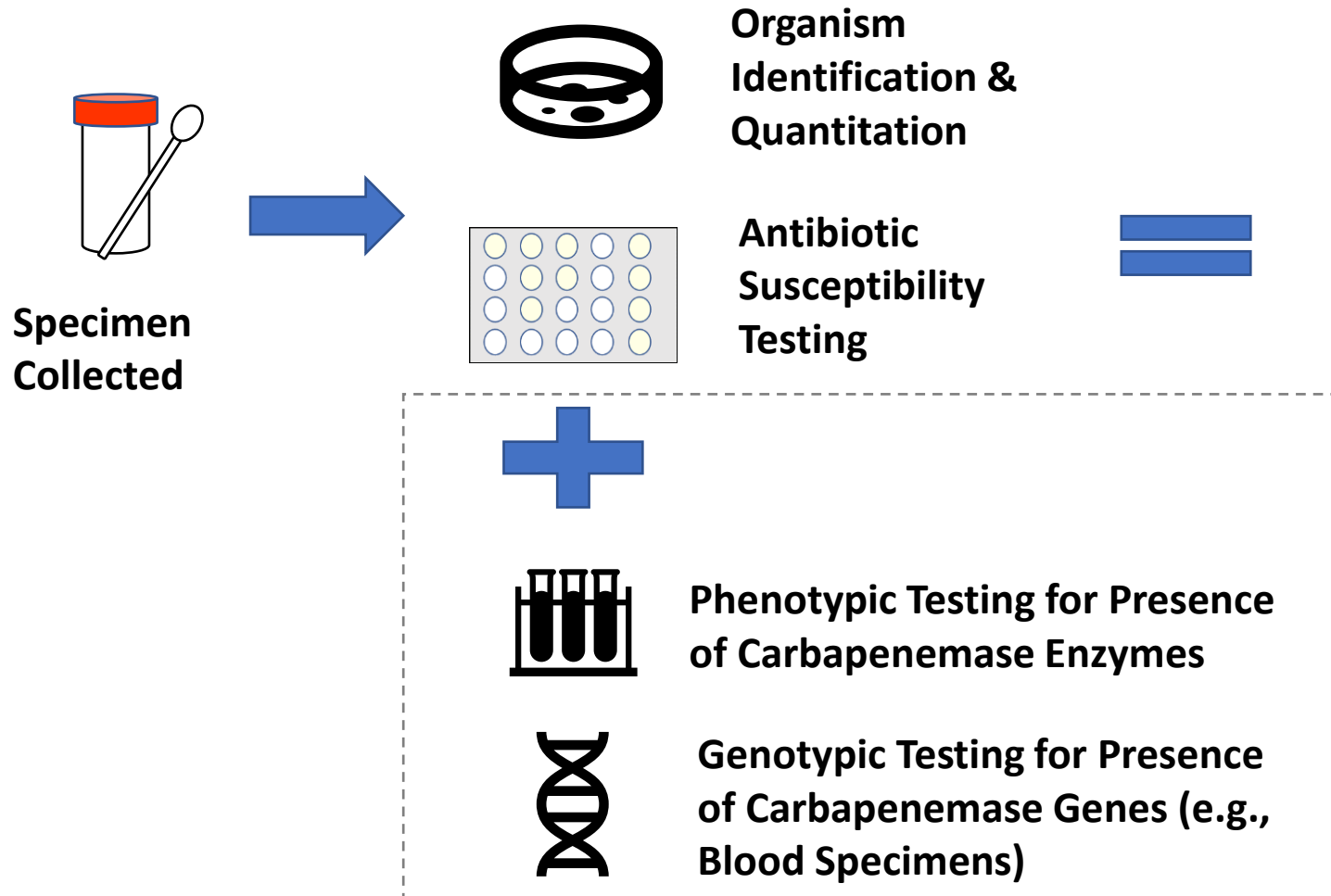
- **Any Enterobacterales** isolate demonstrating carbapenemase production by a phenotypic method
- **Any Enterobacterales** isolate with a known carbapenemase resistance mechanism by a recognized molecular test
- If laboratories are unable to detect CP-CRE (i.e., cannot test for carbapenemase production or carbapenemase resistance mechanism), **any Enterobacterales** isolate with a minimum inhibitory concentration of ≥ 4 mcg/ml for meropenem, imipenem, or doripenem, or ≥ 2 mcg/ml for ertapenem by antimicrobial susceptibility testing
 - *Morganella, Proteus, Providencia* spp. may have intrinsic resistance to imipenem. Only those isolates that are resistant to 1 or more carbapenems other than imipenem should be reported and submitted.

MDHHS Bureau of Laboratories (BOL):

- Confirm organism identification
- Perform mCIM testing
- Perform PCR testing for KPC, NDM, OXA-48, IMP, VIM
 - If mCIM or PCR are positive, antimicrobial susceptibility testing (AST) will be performed



Clinical Microbiology Laboratory Testing

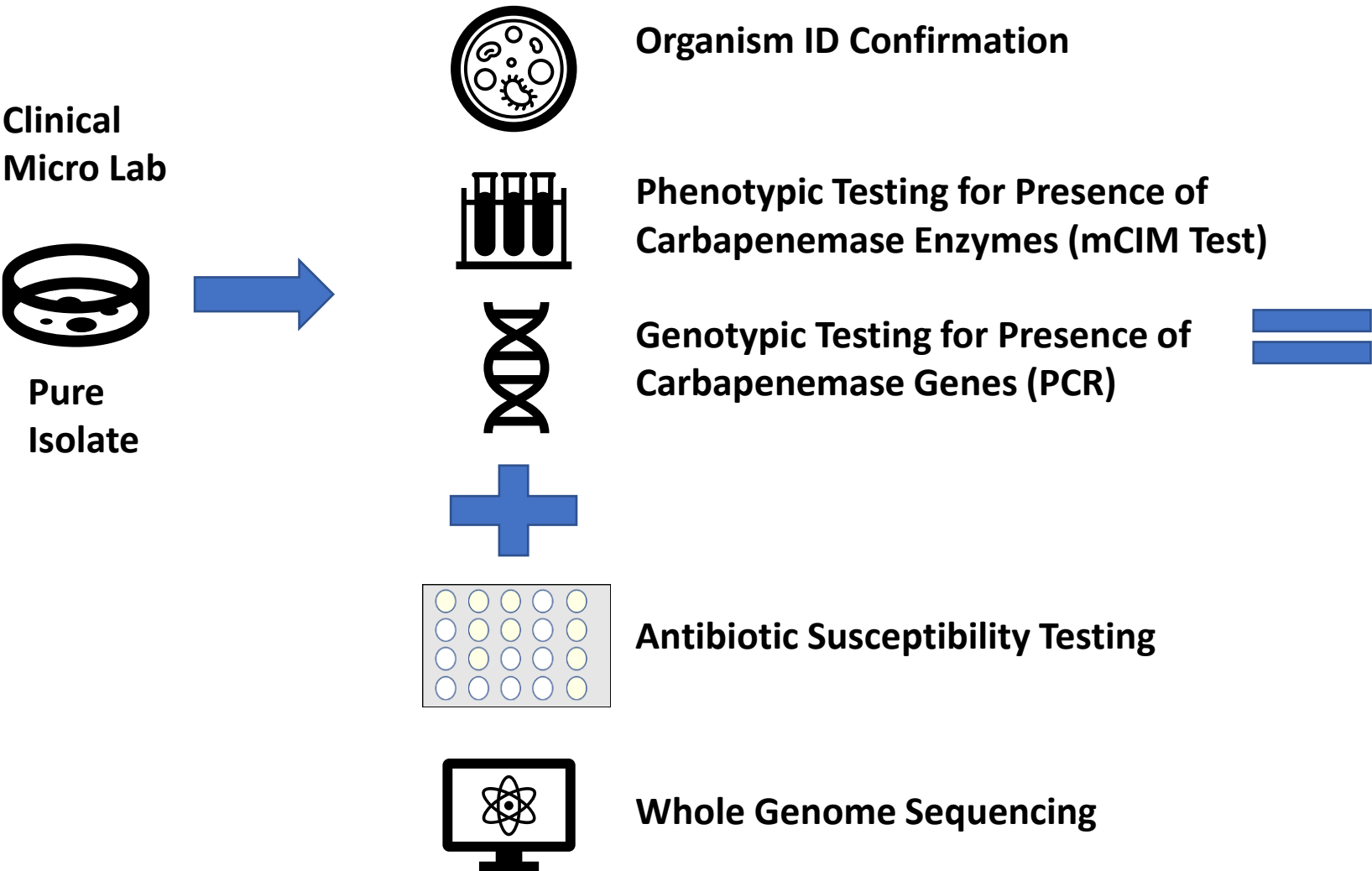


≥100,000 CFU/ml *Klebsiella pneumoniae*, see comment

Comment: Carbapenem resistant Enterobacteriaceae.

<u>Antibiotic</u>	<u>MIC</u>	<u>Interpretation</u>
Ampicillin	≥32	Resistant
Ampicillin/sulbactam	≥32	Resistant
Aztreonam	≥64	Resistant
Cefazolin	≥64	Resistant
Cefepime	2	Resistant
Ceftriaxone	8	Resistant
Ertapenem	2	Resistant
Gentamicin	≤2	Susceptible
Levofloxacin	≤1	Susceptible
Meropenem	1	Susceptible
Piperacillin/tazobactam	64	Intermediate
Tobramycin	≤2	Susceptible
Trimethoprim/sulfamethoxazole	≤2	Susceptible

MDHHS BOL Laboratory Antimicrobial Resistance Confirmation Testing



Antimicrobial Resistance Confirmation (ARC)

Gram Stain
Gram negative bacilli

Culture Results
Confirmed as *Klebsiella pneumoniae*
Identification Performed by MALDI-TOF.

Antimicrobial Susceptibility Results

	<i>Klebsiella pneumoniae</i>	
	MIC - Interpretation	
Amikacin	<=4	S
Aztreonam	>16	R
Cefepime	4	SDD
Cefotaxime	32	R
Ceftazidime	>16	R

Modified Carbapenem Inactivation Method
Positive

Phenotypic test

Modified Carbapenem Inactivation Method (mCIM) screen positive - this isolate demonstrates carbapenemase production. The clinical efficacy of the carbapenems has not been established for treating infections caused by Enterobacteriaceae and Pseudomonas aeruginosa that test carbapenem susceptible but demonstrate carbapenemase production in vitro. ISOLATES THAT ARE mCIM POSITIVE SHOULD BE CONSIDERED RESISTANT TO ALL CARBAPENEMS REGARDLESS OF MIC. MIC REPORTED FOR EPIDEMIOLOGIC PURPOSES ONLY.

PCR Result
KPC (bla-KPC) gene DNA Detected

Molecular test

NDM-1 (bla-NDM-1) gene DNA Not Detected
OXA-48 (bla-OXA-48 like) gene DNA Not Detected
VIM (bla-VIM) gene DNA Not Detected
KPC, NDM, OXA-48, and VIM are the most common carbapenemases in the United States, however there are other less common carbapenemases and other mechanisms of carbapenemase resistance not detected by this PCR assay.

IMP PCR Result
IMP (bla-IMP) gene DNA Not Detected

CP-CRE Case Classification

Confirmed CP-CRE

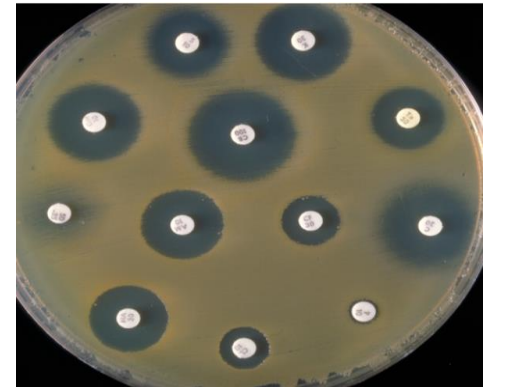
- *Enterobacterales*
 - Positive **phenotypic test** (e.g., mCIM, Carba NP, etc.) OR
 - Positive **molecular test** (e.g., PCR, Cepheid Xpert, etc.) - **carbapenem resistance mechanism**: KPC, NDM, VIM, IMP, OXA-48, etc.

Suspect CP-CRE

- *Enterobacterales*
 - **Resistance to at least 1 carbapenem on susceptibility test- MIC result**
 - No phenotypic or molecular testing done (isolate should be submitted to BOL)

Not a Case

- Negative for phenotypic and molecular tests conducted
- All carbapenems are susceptible (MICs don't match case definition)
- Not *Enterobacterales*



MDHHS BOL ELR Lab Report

Interpretation – Confirmed CP-CRE

Lab Results					
Report Date (mm/dd/yyyy)	Test Name	Reported Test Name/Test Result		Specimen	Collection Date (mm/dd/yyyy)
01/06/2021	Culture Results	Bacteria identified/null	Klebsiella pneumoniae///	Other	12/20/2020
01/06/2021	Antimicrobial Susceptibility Results	Doripenem/null Ertapenem/null Imipenem/null Meropenem/null	///> 2 ///> 4 ///> 8 ///> 8		12/20/2020
01/06/2021	Modified Carbapenem Inactivation Method	Carbapenemase/null	Positive///		12/20/2020
01/06/2021	PCR Result	bla(KPC) gene/null Bacterial carbapenem resistance blaNDM gene/null Bacterial carbapenem resistance blaOXA-48-like gene/null Bacterial carbapenem resistance blaVIM gene/null	KPC (bla-KPC) gene DNA Not Detected/// NDM-1 (bla-NDM-1) gene DNA Detected/// OXA-48 (bla-OXA-48 like) gene DNA Not Detected/// VIM (bla-VIM) gene DNA Not Detected///		12/20/2020
01/06/2021	IMP PCR Result	Bacterial carbapenem resistance blaIMP gene/null	IMP (bla-IMP) gene DNA Not Detected///		12/20/2020
01/06/2021	Carbapenem resistance genes	Carbapenem resistance genes/ARC	Klebsiella pneumoniae///		12/20/2020
01/05/2021	Culture Results	Bacteria identified/	Klebsiella pneumoniae///	Other	12/20/2020
01/05/2021	PCR Result	bla(KPC) gene/ Bacterial carbapenem resistance blaNDM gene/ Bacterial carbapenem resistance blaOXA-48-like gene/ Bacterial carbapenem resistance blaVIM gene/	KPC (bla-KPC) gene DNA Not Detected/// NDM-1 (bla-NDM-1) gene DNA Detected/// OXA-48 (bla-OXA-48 like) gene DNA Not Detected/// VIM (bla-VIM) gene DNA Not Detected///		12/20/2020

Antimicrobial Resistance Confirmation (ARC)

Gram Stain

Gram negative bacilli

Culture Results

Confirmed Identification by MALDI-TOF - Klebsiella pneumoniae

Antimicrobial Susceptibility Results

	Klebsiella pneumoniae	
	MIC - Interpretation	
Aztreonam	>16	R
Cefepime	>16	R

Modified Carbapenem Inactivation Method

Positive

Modified Carbapenem Inactivation Method (mCIM) screen positive - this isolate demonstrates carbapenemase production. The clinical efficacy of the carbapenems has not been established for treating infections caused by Enterobacteriaceae and Pseudomonas aeruginosa that test carbapenem susceptible but demonstrate carbapenemase production in vitro. ISOLATES THAT ARE mCIM POSITIVE SHOULD BE CONSIDERED RESISTANT TO ALL CARBAPENEMS REGARDLESS OF MIC. MIC REPORTED FOR EPIDEMIOLOGIC PURPOSES ONLY.

PCR Result

KPC (bla-KPC) gene DNA Not Detected

NDM-1 (bla-NDM-1) gene DNA Detected

IMP PCR Result

IMP (bla-IMP) gene DNA Not Detected

16S rRNA Sequencing, PCR, and MALDI-TOF tests were developed and their performance characteristics determined by the Michigan Department of Health and Human Services (MDHHS). They have not been cleared or approved by the U.S. Food and Drug Administration (FDA). The FDA has determined that such clearance or approval is not necessary if performance characteristics are verified at the testing laboratory.

Initial screening for carbapenemase genes performed using Cepheid GeneXpert which has been FDA approved for this testing.

MDHHS BOL ELR Lab Report Interpretation – Not a Case, CP-CRE

Date Collected	07/22/2021	Patient Last Name	[REDACTED]
Time Collected	1014	Patient First Name	[REDACTED]
Date Received	07/29/2021	Patient DOB	[REDACTED]
Specimen Type	SPUTUM	Submitter Patient ID	[REDACTED]
		Gender	[REDACTED]
		Physician	[REDACTED]
		Submitter Identifier	P51690
		Reason for Test	DIAGNOSIS

TEST RESULTS

Antimicrobial Resistance Confirmation (ARC)

Gram Stain

Direct Gram Stain Not Done

Culture Results

Confirmed Identification by MALDI-TOF - Enterobacter cloacae complex

Modified Carbapenem Inactivation Method

Negative

Modified Carbapenem Inactivation Method (mCIM) screen negative - not all carbapenemase-producing isolates of Enterobacteriaceae and Pseudomonas aeruginosa are mCIM positive.

16S rRNA Sequencing, PCR, and MALDI-TOF tests were developed and their performance characteristics determined by the Michigan Department of Health and Human Services (MDHHS). They have not been cleared or approved by the U.S. Food and Drug Administration (FDA). The FDA has determined that such clearance or approval is not necessary if performance characteristics are verified at the testing laboratory.

Initial screening for carbapenemase genes performed using Cepheid GeneXpert which has been FDA approved for this testing.

Lab Reports						Help
Date Received	Collection Date	Test Name (* Case Associated)	Result	Electronic		
08/11/2021	07/22/2021	Culture Results	Enterobacter cloacae complex	Yes	View	
08/11/2021	07/22/2021	Modified Carbapenem Inactivation Method	Negative	Yes	View	
08/11/2021	07/22/2021	Carbapenem resistance genes	Enterobacter cloacae complex	Yes	View	
		Modified Carbapenem Inactivation				

manual case entry

⚠ Urine Culture (Source: Clean Catch)

Collected: 12/23/2020 15:11 Status: Final result Visible to patient: No (not released)

Specimen Information: Urine, Indwelling Catheter
Indwelling catheter

Component

Special Requests None

Results >100000 CFU per ml Klebsiella pneumoniae!

Susceptibility

Klebsiella pneumoniae

Antibiotic	Interpretation	Value	Method
Amikacin*	Susceptible	≤2	MIC MYLA
Ampicillin	Resistant	≥32	MIC MYLA
Ampicillin/Sulbactam*	Resistant	≥32	MIC MYLA
Cefazolin	Resistant	≥64	MIC MYLA
Cefepime	Resistant	≤1	MIC MYLA
Ceftriaxone	Resistant	≤1	MIC MYLA
Ciprofloxacin	Susceptible	≤0.25	MIC MYLA
Ertapenem	Resistant	≤0.5	MIC MYLA
Gentamicin	Susceptible	≤1	MIC MYLA
Meropenem	Resistant	≥16	MIC MYLA
Meropenem		This organism	MIC MYLA

This organism is resistant to one or more carbapenems. Follow infection control requirements for CRE.

Laboratory Testing and Microbiology Information			
Type of facility where specimen was collected: <div style="display: flex; justify-content: space-between; margin-top: 5px;"> <input checked="" type="radio"/> Acute Care Hospital <input type="radio"/> Long-Term Acute Care Hospital <input type="radio"/> Long-Term Care Facility <input type="radio"/> Outpatient <input type="radio"/> Other </div> <div style="display: flex; justify-content: space-between; margin-top: 5px;"> <input type="radio"/> Autopsy <input type="radio"/> Unknown </div>			
Date Specimen Collected (mm/dd/yyyy) County of the facility where specimen collected: Facility where specimen collected: <div style="display: flex; justify-content: space-between; margin-top: 5px;"> <input style="width: 20%;" type="text" value="12/23/2020"/> <input style="width: 20%; background-color: black; color: black;" type="text"/> <input style="width: 60%; background-color: black; color: black;" type="text"/> </div>			
Clinical Specimen Source: Other source, specify: Specimen site, if available: <div style="display: flex; justify-content: space-between; margin-top: 5px;"> <input style="width: 25%;" type="text" value="Urine specimen"/> <input style="width: 35%; background-color: black; color: black;" type="text"/> <input style="width: 40%; background-color: black; color: black;" type="text"/> </div>			
Organism: <input type="radio"/> Other, specify: <div style="display: flex; justify-content: space-between; margin-top: 5px;"> <input style="width: 30%;" type="text" value="Klebsiella pneumoniae"/> <input style="width: 65%; background-color: black; color: black;" type="text"/> </div>			
Was Antimicrobial Susceptibility Testing performed? <input type="radio"/> Yes <input type="radio"/> No <input type="radio"/> Unknown			
Antimicrobial Susceptibility Testing Results:			
Antimicrobial	Minimum Inhibitory Concentration (MIC) (ug/ml)	Interpretation (S, susceptible; I, Intermediate; R, resistant)	
Doripenem	<input style="width: 90%;" type="text"/>	<input style="width: 90%;" type="text"/>	
Ertapenem	<input style="width: 90%;" type="text"/>	<input style="width: 90%;" type="text"/>	
Imipenem	<input style="width: 90%;" type="text"/>	<input style="width: 90%;" type="text"/>	
Meropenem	<input style="width: 90%;" type="text" value="≥16"/>	<input style="width: 90%;" type="text" value="R"/>	
Phenotype Tests: If Other, specify: Result: <div style="display: flex; justify-content: space-between; margin-top: 5px;"> <input style="width: 35%;" type="text" value="Not Tested"/> <input style="width: 30%; background-color: black; color: black;" type="text"/> <div> <input type="radio"/> Positive <input type="radio"/> Negative <input type="radio"/> Indeterminate </div> </div>			
Molecular Tests: If Other, specify: Result: <div style="display: flex; justify-content: space-between; margin-top: 5px;"> <input style="width: 35%;" type="text" value="Not Tested"/> <input style="width: 30%; background-color: black; color: black;" type="text"/> <div> <input type="radio"/> Positive <input type="radio"/> Negative <input type="radio"/> Indeterminate </div> </div>			
Resistance Mechanism for Carbapenemase Testing	Response		
KPC	<input type="radio"/> Detected <input type="radio"/> Not detected <input type="radio"/> Not tested <input type="radio"/> Invalid		
NDM	<input type="radio"/> Detected <input type="radio"/> Not detected <input type="radio"/> Not tested <input type="radio"/> Invalid		
VIM	<input type="radio"/> Detected <input type="radio"/> Not detected <input type="radio"/> Not tested <input type="radio"/> Invalid		
IMP	<input type="radio"/> Detected <input type="radio"/> Not detected <input type="radio"/> Not tested <input type="radio"/> Invalid		
OXA-48	<input type="radio"/> Detected <input type="radio"/> Not detected <input type="radio"/> Not tested <input type="radio"/> Invalid		
OXA-23	<input type="radio"/> Detected <input type="radio"/> Not detected <input type="radio"/> Not tested <input type="radio"/> Invalid		
Other, specify <input style="width: 30%;" type="text"/>	<input type="radio"/> Detected <input type="radio"/> Not detected <input type="radio"/> Not tested <input type="radio"/> Invalid		
Clinical Lab Specimen ID (unique isolate No.): Bureau of Labs Specimen ID: WGS Accession ID: <div style="display: flex; justify-content: space-between; margin-top: 5px;"> <input style="width: 30%; background-color: black; color: black;" type="text"/> <input style="width: 30%; background-color: black; color: black;" type="text"/> <input style="width: 40%; background-color: black; color: black;" type="text"/> </div>			

Clinical Lab ELR Report Interpretation – Suspect CP-CRE

Lab Report Date (mm/dd/yyyy) :	05/23/2021
Ordering Provider	
First :	ELI LEON
Affiliation :	BEAUMONT HEALTH SYSTEM
Street :	17000 HUBBARD DR
City :	DEARBORN
County :	Wayne
State :	Michigan
Zip :	48126
Phone number :	313-982-4351
Ext :	
Laboratory Information	
Lab Name* :	Oakwood Hospital - Dearborn
Street :	18101 Oakwood Blvd
City :	Dearborn
County :	Wayne
State :	Michigan
Zip :	48124
Phone number :	
Specimen Information	
Specimen Collection Date (mm/dd/yyyy) :	05/19/2021
Specimen Source :	
Specimen Site :	
Specimen Site Text :	
Specimen ID :	
Results	
Reported Test Name : ceFAZolin Islt MIC/null	
Numeric Result :	>= 64
Abnormal Flags/Susceptibility Results:	R
Reported Test Name : Cefepime Islt MIC/null	
Numeric Result :	<= 1
Abnormal Flags/Susceptibility Results:	R
Reported Test Name : cefTRIAxone Islt MIC/null	
Numeric Result :	<= 1
Abnormal Flags/Susceptibility Results:	R
Reported Test Name : Ertapenem Islt MIC/null	
Text Result :	2
Numeric Result :	
Abnormal Flags/Susceptibility Results:	R

Lab Order Information	
Test Name* :	Bacteria Ur Cult
Lab Report Date (mm/dd/yyyy) :	05/23/2021
Ordering Provider	
First :	
Last :	
Affiliation :	
Street :	17000 HUBBARD DR
City :	
County :	
State :	
Zip :	
Phone number :	
Ext :	
Laboratory Information	
Lab Name* :	Oakwood Hospital - Dearborn
Street :	18101 Oakwood Blvd
City :	Dearborn
County :	Wayne
State :	Michigan
Zip :	48124
Phone number :	
Specimen Information	
Specimen Collection Date (mm/dd/yyyy) :	05/19/2021
Specimen Source :	Urine specimen
Specimen Site :	
Specimen Site Text :	URINE SPECIMEN OBTAINED BY CLEAN CATCH PROCED
Specimen ID :	
Results	
Reported Test Name : Bacteria Ur Cult/Culture, Urine	
Coded Result :	CARBAPENEM RESISTANT ENTEROBACTER CLOACAE COMPLEX
Numeric Result :	
Abnormal Flags/Susceptibility Results:	A
Comments :	>100,000 CFU/ml Enterobacter cloacae complex, (CRE) MDR - This isolate is resistant to a carbapenem(s)(CRE). Initiate contact precautions. Consider Infectious Diseases consult.

Does this Isolate Meet Reporting Requirements?

- ✓ *Klebsiella pneumoniae*
- ✓ Carbapenemase production
- ✓ KPC carbapenemase gene detected
- ✓ Ertapenem MIC ≥ 4
- ✓ Meropenem MIC ≥ 16

= Confirmed CP-CRE Case

$\geq 100,000$ CFU/ml *Klebsiella pneumoniae*, see comment

Comment: Carbapenem resistant Enterobacteriaceae. Carbapenemase producer. KPC detected.

<u>Antibiotic</u>	<u>MIC</u>	<u>Interpretation</u>
Ampicillin	≥ 32	Resistant
Ampicillin/sulbactam	≥ 32	Resistant
Aztreonam	≥ 64	Resistant
Cefazolin	≥ 64	Resistant
Cefepime	2	Resistant
Ceftriaxone	8	Resistant
Ertapenem	≥ 4	Resistant
Gentamicin	≤ 2	Sensitive
Levofloxacin	≤ 1	Sensitive
Meropenem	≥ 16	Resistant
Piperacillin/tazobactam	64	Intermediate
Tobramycin	≤ 2	Sensitive
Trimethoprim/sulfamethoxazole	≤ 2	Sensitive

Does this Isolate Meet Reporting Requirements?

✓ *Enterobacter cloacae*

X No phenotypic or molecular carbapenemase testing reported

✓ Ertapenem MIC = 2

X Meropenem MIC = 1

= Suspect CP-CRE Case

≥100,000 CFU/ml *Enterobacter cloacae*, see comment

Comment: Carbapenem resistant Enterobacteriaceae.

<u>Antibiotic</u>	<u>MIC</u>	<u>Interpretation</u>
Ampicillin	≥32	Resistant
Ampicillin/sulbactam	≥32	Resistant
Aztreonam	≥64	Resistant
Cefazolin	≥64	Resistant
Cefepime	2	Resistant
Ceftriaxone	8	Resistant
Ertapenem	2	Resistant
Gentamicin	≤2	Susceptible
Levofloxacin	≤1	Susceptible
Meropenem	1	Susceptible
Piperacillin/tazobactam	64	Intermediate
Tobramycin	≤2	Susceptible
Trimethoprim/sulfamethoxazole	≤2	Susceptible

Does this Isolate Meet Reporting Requirements?

✓ *Escherichia coli*

X No phenotypic or molecular carbapenemase testing reported

? Ertapenem and meropenem reported as 'Resistant' but no MIC value reported

= Can not tell if it meets the case definition or not

≥100,000 CFU/ml *Escherichia coli*, see comment

Comment: Carbapenem resistant Enterobacteriaceae.

<u>Antibiotic</u>	<u>MIC</u>	<u>Interpretation</u>
Ampicillin	≥32	Resistant
Ampicillin/sulbactam	≥32	Resistant
Aztreonam		Resistant
Cefazolin		Resistant
Cefepime		Resistant
Ceftriaxone		Resistant
Ertapenem		Resistant
Gentamicin	≤2	Sensitive
Levofloxacin	≤1	Sensitive
Meropenem		Resistant
Piperacillin/tazobactam	64	Intermediate
Tobramycin	≤2	Sensitive
Trimethoprim/sulfamethoxazole	≤2	Sensitive

Tips for CP-CRE Reporting

- **Confirm the organism identification**
 - Enterobacterale - Enterobacterales is an order of different types of bacteria which include *Escherichia*, *Klebsiella*, *Enterobacter*, *Salmonella*, *Shigella*, *Citrobacter* and *Yersinia*.
- **Review carbapenem Susceptibility testing MIC values**
 - Doripenem, imipenem, or meropenem ≥ 4 $\mu\text{g/ml}$; or ertapenem ≥ 2 $\mu\text{g/ml}$
 - If there are no MIC values reported (e.g., “Resistant”) or no carbapenems reported in MDSS, call the laboratory and ask to speak to a bench technologist
- **Check for phenotypic carbapenemase testing**
 - ‘Carbapenemase detected’ or ‘Carbapenemase not detected’
 - Confirm the method used: mCIM, CarbaNP, MBL test
- **Check for molecular carbapenemase testing for resistance mechanisms**
 - KPC, NDM, OXA-48, VIM, IMP



Case Investigation Forms

“Case Report Form (CRF)” or “Case Detail Form”, or “Case Investigation Form”

- Sections
 - Investigation Information
 - Patient Information
 - Demographics
 - Referral Information
 - Laboratory Testing and Microbiology Information
 - Clinical Information
 - Other Information
 - Case Notes
 - Lab Results

Back

Print

CP-CRE Case Report

Carbapenemase-Producing Carbapenem-Resistant Enterobacteriaceae (CP-CRE)

Michigan Department of Health and Human Services

Communicable Disease Division

Back

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Candida auris Case Report

Michigan Department of Health and Human Services

Communicable Disease Division

Expand all

Collapse all

Investigation Information


Investigation ID	Onset Date (mm/dd/yyyy)	Diagnosis Date (mm/dd/yyyy)	Referral Date (mm/dd/yyyy)	Case Entry Date (mm/dd/yyyy)
	<input type="text"/>	<input type="text"/>	<input type="text"/>	<input type="text"/>
Investigation Status Active	Case Status <input type="radio"/> Confirmed <input type="radio"/> Probable			<input type="radio"/> Confirmed - Non Resident <input type="radio"/> Suspect <input type="radio"/> Not a Case <input type="radio"/> Non-Michigan Case <input type="checkbox"/> State Prison Case
Patient Status Alive	Patient Status Date (mm/dd/yyyy)	Case Disposition	Case Updated Date (mm/dd/yyyy)	Case Completion Date (mm/dd/yyyy)
	<input type="text"/>	<input type="text"/>	<input type="text"/>	<input type="text"/>
Date of Death (mm/dd/yyyy)	Investigator First Name: Last Name:		Part of an outbreak?	Outbreak Name
<input type="text"/>	<input type="text"/>		<input type="text"/>	<input type="text"/>
Case Type: <input type="radio"/> Clinical Case <input type="radio"/> Colonization/Screening Case				
Clinical Candida auris Case Only: Was patient previously counted as a colonization/screening case? <input type="radio"/> Yes <input type="radio"/> No <input type="radio"/> Unknown If patient was previously counted as a colonization/screening case, please provide the related case ID(s) <input type="text"/>				

Patient Information

Patient ID	First	Last	Middle
<input type="text"/>	<input type="text"/>	<input type="text"/>	<input type="text"/>
Street Address			
<input type="text"/>			

Collapse all

al Date (mm/dd/yyyy)	Case Entry Date (mm/dd/yyyy)
<input type="text"/>	<input type="text"/>
a Case -Michigan Case	<input type="checkbox"/> State Prison Case
se Updated Date (mm/dd/yyyy)	Case Completion Date (mm/dd/yyyy)
<input type="text"/>	<input type="text"/>
rt of an outbreak?	Outbreak Name
<input type="text"/>	<input type="text"/>
Unknown	
<input type="text"/>	
Middle	
<input type="text"/>	<input type="text"/>



Candida auris Laboratory Testing

- Laboratory Testing information is required to determine case classification

- Date collected
- Specimen source
- Test Type



[Back](#) [Print](#)

Candida auris Case Report

Michigan Department of Health and Human Services

Communicable Disease Division

+

Demographics

+

Referral Information

-

Laboratory Testing and Microbiology Information

Type of facility where specimen was collected:

☐ Acute Care Hospital

☐ Long-Term Acute Care Hospital

☐ Long-Term Care Facility

☒ Outpatient

☐ Other

☐ Autopsy

☐ Unknown

Date Specimen Collected (mm/dd/yyyy)

County of the facility where specimen collected:

Facility where specimen collected:

05/20/2021

Oakland

For Clinical Case:

Specimen Source:

Other source, specify:

Other

Ear Drainage

For Colonization/Screening Case:

Screening swab anatomical site:

Other site:

Clinical Lab Specimen ID (unique isolate No.):

Bureau of Labs Specimen ID:

WGS Accession ID:

Test Type:

Test Method (manufacturer/brand, type of PCR, etc.):

Result:

MALDI-TOF

Bruker

☒ Detected

☐ Not Detected

☐ Indeterminate

Other test, specify:

Test Type:

Test Method (manufacturer/brand, type of PCR, etc.):

Result:

☐ Detected

☐ Not Detected

☐ Indeterminate

Other test, specify:

Test Type:

Test Method (manufacturer/brand, type of PCR, etc.):

Result:

☐ Detected

☐ Not Detected

☐ Indeterminate

Other test, specify:

CP-CRE Laboratory Testing

- **Laboratory Testing information is required to determine case classification**

- Date collected
- Specimen source
- Organism
- **Susceptibility test: MIC**
 - need actual numerical value
- **Phenotype test: Carbapenemase testing**
 - e.g., mCIM, CarbaNP
- **Molecular test: Resistance mechanism-gene testing**
 - e.g. PCR, Carba-R



Organism: <input type="radio"/> <i>Klebsiella aerogenes</i> <input type="radio"/> <i>Klebsiella pneumoniae</i> <input type="radio"/> <i>Enterobacter cloacae</i> <input type="radio"/> <i>Escherichia coli</i> <input type="radio"/> Other, specify: <input type="text"/>		
<input type="radio"/> <i>Klebsiella oxytoca</i> <input type="radio"/> <i>Klebsiella, other</i> <input type="radio"/> <i>Enterobacter, other</i> <input type="text"/>		
Was Antimicrobial Susceptibility Testing performed? <input type="radio"/> Yes <input type="radio"/> No <input type="radio"/> Unknown		
Antimicrobial Susceptibility Testing Results:		
Antimicrobial	Minimum Inhibitory Concentration (MIC) (ug/ml)	Interpretation (S, susceptible; I, Intermediate; R, resistant)
Doripenem	<input type="text"/>	<input type="text"/>
Ertapenem	<input type="text"/>	<input type="text"/>
Imipenem	<input type="text"/>	<input type="text"/>
Meropenem	<input type="text"/>	<input type="text"/>
Phenotype Tests: <input type="text"/> If Other, specify: <input type="text"/> Result: <input type="radio"/> Positive <input type="radio"/> Negative <input type="radio"/> Indeterminate		
Molecular Tests: <input type="text"/> If Other, specify: <input type="text"/> Result: <input type="radio"/> Positive <input type="radio"/> Negative <input type="radio"/> Indeterminate		
Resistance Mechanism for Carbapenemase Testing		Response
KPC	<input type="radio"/> Detected <input type="radio"/> Not detected <input type="radio"/> Not tested <input type="radio"/> Invalid	
NDM	<input type="radio"/> Detected <input type="radio"/> Not detected <input type="radio"/> Not tested <input type="radio"/> Invalid	
VIM	<input type="radio"/> Detected <input type="radio"/> Not detected <input type="radio"/> Not tested <input type="radio"/> Invalid	
IMP	<input type="radio"/> Detected <input type="radio"/> Not detected <input type="radio"/> Not tested <input type="radio"/> Invalid	
OXA-48	<input type="radio"/> Detected <input type="radio"/> Not detected <input type="radio"/> Not tested <input type="radio"/> Invalid	
OXA-23	<input type="radio"/> Detected <input type="radio"/> Not detected <input type="radio"/> Not tested <input type="radio"/> Invalid	
Other, specify <input type="text"/>	<input type="radio"/> Detected <input type="radio"/> Not detected <input type="radio"/> Not tested <input type="radio"/> Invalid	
Clinical Lab Specimen ID (unique isolate No.): <input type="text"/>	Bureau of Labs Specimen ID: <input type="text"/>	WGS Accession ID: <input type="text"/>

Clinical Info for CP-CRE and *Candida auris*

- Healthcare exposures
 - Acute care, long-term care
- Travel
 - Location
 - Healthcare abroad
 - Very important for confirmed NDM, OXA-48, IMP, or VIM cases

Travel History			
Does the patient have a history of Travel outside the USA in the last 12 months? <input type="radio"/> Yes <input type="radio"/> No <input type="radio"/> Unknown			
If Yes, please indicate the country and dates of travel			
Country:	Dates:	From	To
<input type="text"/>	(mm/dd/yyyy)	<input type="text"/>	<input type="text"/>
Country:	Dates:	From	To
<input type="text"/>	(mm/dd/yyyy)	<input type="text"/>	<input type="text"/>
Country:	Dates:	From	To
<input type="text"/>	(mm/dd/yyyy)	<input type="text"/>	<input type="text"/>
Has the patient received Healthcare outside the USA within the last 12 months? <input type="radio"/> Yes <input type="radio"/> No <input type="radio"/> Unknown			
If Yes, what Country: <input type="text"/>			
Has the patient received Healthcare outside the state of Michigan within the last 12 months? <input type="radio"/> Yes <input type="radio"/> No <input type="radio"/> Unknown			
If Yes, what State: <input type="text"/>			

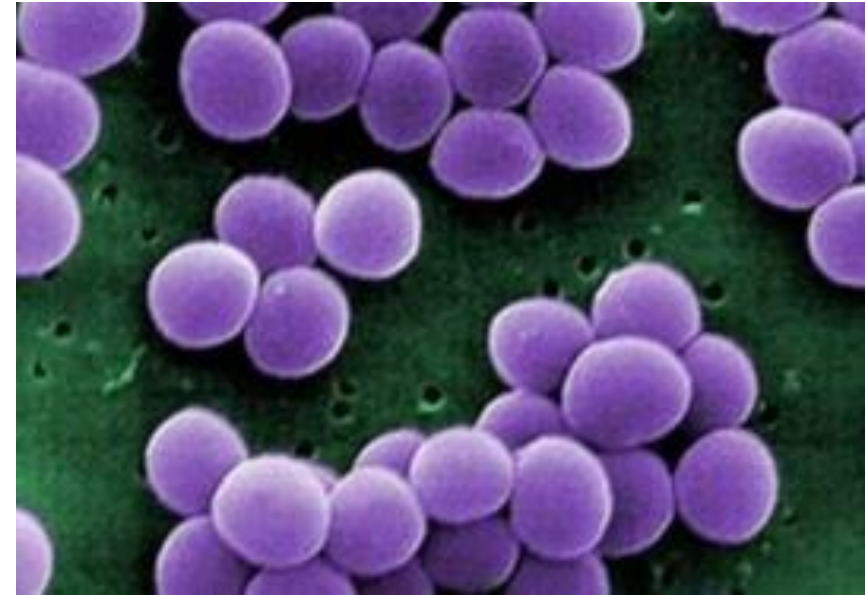
Clinical Information			
Date of Patient Admission or Presentation (mm/dd/yyyy)		Date Patient was placed in Contact Precautions/Isolation (if an inpatient) (mm/dd/yyyy)	
<input type="text"/>		<input type="text"/>	
Patient Admitted/Presented From:			
<input type="radio"/> Long-Term Care/Skilled Nursing Facility		<input type="radio"/> Outside Acute Care Hospital	
<input type="radio"/> Long-Term Acute Care Hospital		<input type="radio"/> Unknown	
<input type="radio"/> Home		<input type="radio"/> Other, specify <input type="text"/>	
Date of Patient Discharge (mm/dd/yyyy)		Was information on CRE status shared with transferring agency and admitting facility:	
<input type="text"/>		<input type="radio"/> Yes <input type="radio"/> No <input type="radio"/> Unknown	
Patient Discharged to:			
<input type="radio"/> Long-Term Care/Skilled Nursing Facility		<input type="radio"/> Outside Acute Care Hospital	
<input type="radio"/> Long-Term Acute Care Hospital		<input type="radio"/> Unknown	
<input type="radio"/> Home		<input type="radio"/> Other, specify <input type="text"/>	
Has Patient previously been hospitalized in an Acute Care Hospital in the last 90 days: <input type="radio"/> Yes <input type="radio"/> No <input type="radio"/> Unknown			
If Yes, please indicate the facility name and dates of stay (if known)			
Facility:	Dates:	From	To
<input type="text"/>	(mm/dd/yyyy)	<input type="text"/>	<input type="text"/>
Has Patient been admitted to a Long-Term Acute Care Hospital in the last 90 days: <input type="radio"/> Yes <input type="radio"/> No <input type="radio"/> Unknown			
If Yes, please indicate the facility name and dates of stay (if known)			
Facility:	Dates:	From	To
<input type="text"/>	(mm/dd/yyyy)	<input type="text"/>	<input type="text"/>
Has Patient been admitted to a Long-Term Care Facility (e.g., nursing home, SNF) in the last 90 days: <input type="radio"/> Yes <input type="radio"/> No <input type="radio"/> Unknown			
If Yes, please indicate the facility name and dates of stay (if known)			
Facility:	Dates:	From	To
<input type="text"/>	(mm/dd/yyyy)	<input type="text"/>	<input type="text"/>
Indwelling Devices (in place within 2 calendar days of specimen collection):			
Central Venous Line: <input type="radio"/> Yes <input type="radio"/> No <input type="radio"/> Unknown		Mechanical Ventilation: <input type="radio"/> Yes <input type="radio"/> No <input type="radio"/> Unknown	
Urinary Catheter: <input type="radio"/> Yes <input type="radio"/> No <input type="radio"/> Unknown		Wound VAC (vacuum-assisted closure): <input type="radio"/> Yes <input type="radio"/> No <input type="radio"/> Unknown	

VISA and VRSA

Vancomycin-intermediate *Staphylococcus aureus* (VISA)

Vancomycin-resistant *Staphylococcus aureus* (VRSA)

- Vancomycin is a critical antibiotic for the treatment of MRSA
- Requires a rapid and aggressive containment response
- 16 U.S. cases to date
 - Primarily in MI and DE
 - Last identified in 2021 in MI & NC (first cases since 2015!)



Vancomycin-resistant *Staphylococcus aureus* (VRSA)

- Thought to result from MRSA containing a pSK41-type plasmid and VRE containing *vanA* encoded on an Inc18-like plasmid
 - All 16 cases were *vanA* +
- Classified based on minimum inhibitory concentration (MIC) on susceptibility test

Vancomycin-susceptible *S. aureus* (VSSA)

- Vancomycin MIC ≤ 2 $\mu\text{g/ml}$

Vancomycin-intermediate *S. aureus* (VISA)

- Vancomycin MIC = 4-8 $\mu\text{g/ml}$.

Vancomycin-resistant *S. aureus* (VRSA)

- Vancomycin MIC ≥ 16 $\mu\text{g/ml}$.

Note: The breakpoints for S. aureus and vancomycin differ from those for other Staphylococcus species. (2015 CLSI M100-S25).

Lab Reports						Help
Date Received	Collection Date	Test Name (* Case Associated)	Result	Electronic		
06/21/2021	06/09/2021	Antimicrobial Susceptibility Results *	> 128	Yes	View	
06/21/2021	06/09/2021	vanA PCR Result *	vanA gene Detected	Yes	View	
06/21/2021	06/09/2021	Bacteria identified *	vanA gene Detected	Yes	View	

MDSS VRSA Case Investigation

- Report requires extensive case information
- Reports of suspected VRSA cases
 - Often mixed cultures of VRE and MRSA:
 - Ask laboratories to re-streak for purity and repeat AST
 - *S. aureus* isolates with vancomycin MICs ≥ 4 $\mu\text{g/ml}$ should be confirmed by a validated method and infection control should be notified
 - *S. aureus* isolates with a vancomycin MICs of ≥ 8 $\mu\text{g/ml}$ should be submitted to health departments and/or CDC for confirmation by a reference method
 - Notify health departments
- Ask facilities to save any MRSA and VRE isolates
- Patients with suspected VRSA should be place in isolation and contact precautions while awaiting results

Back

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Vancomycin-Resistant Staphylococcus aureus (VRSA)

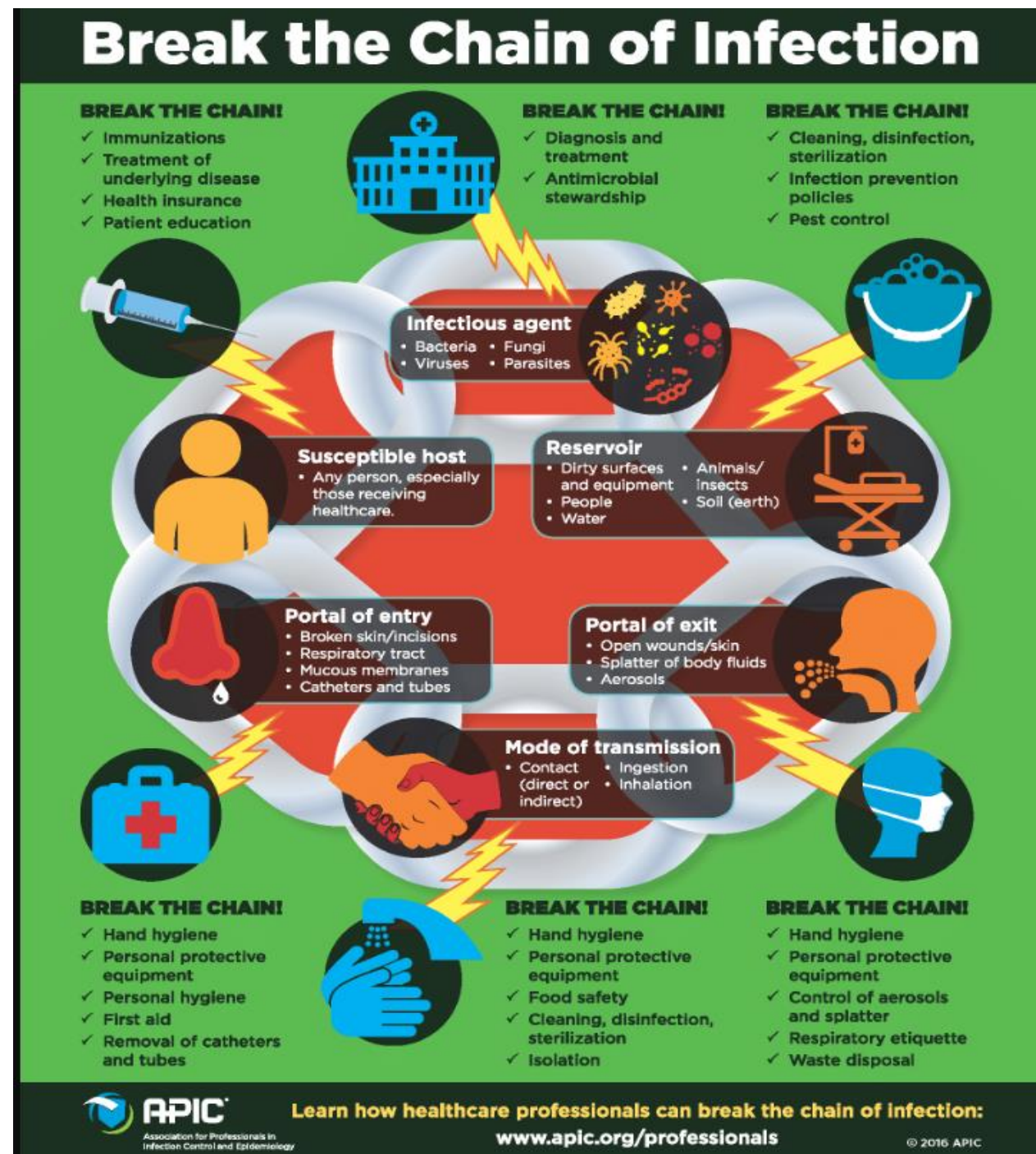
Michigan Department of Health and Human Services

Communicable Disease Division

[Expand all](#)[Collapse all](#)

+	Investigation Information
+	Patient Information
+	Demographics
+	Referral Information
+	Referral Information Continued
+	Facility Information (at time of referral)
+	Isolate Information
+	Clinical Information
+	Clinical Information cont.
+	Clinical Information cont.
+	Other Information
+	Case Notes

Containment of Targeted MDROs

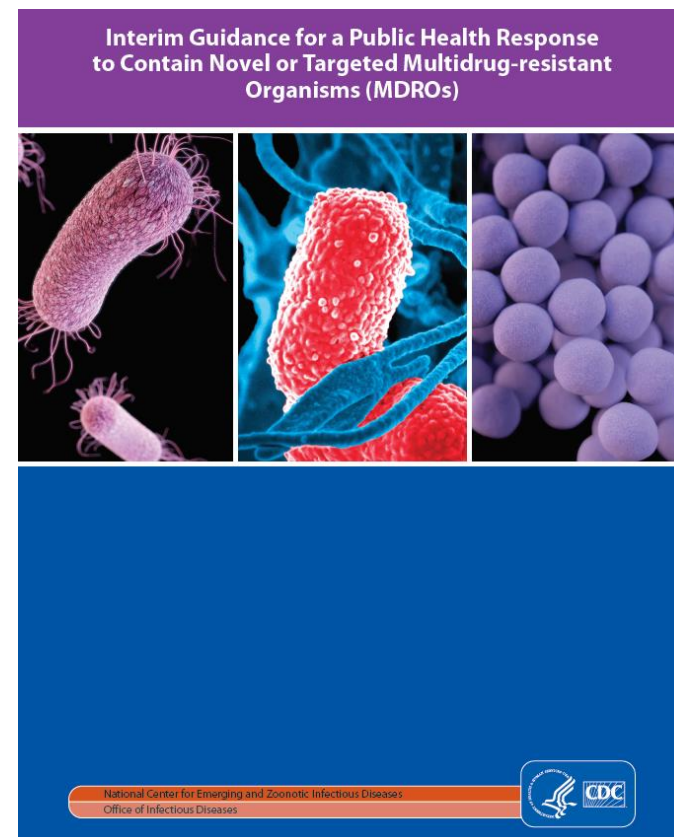


2022 Update: Containment of Targeted MDROs

COMING
SOON

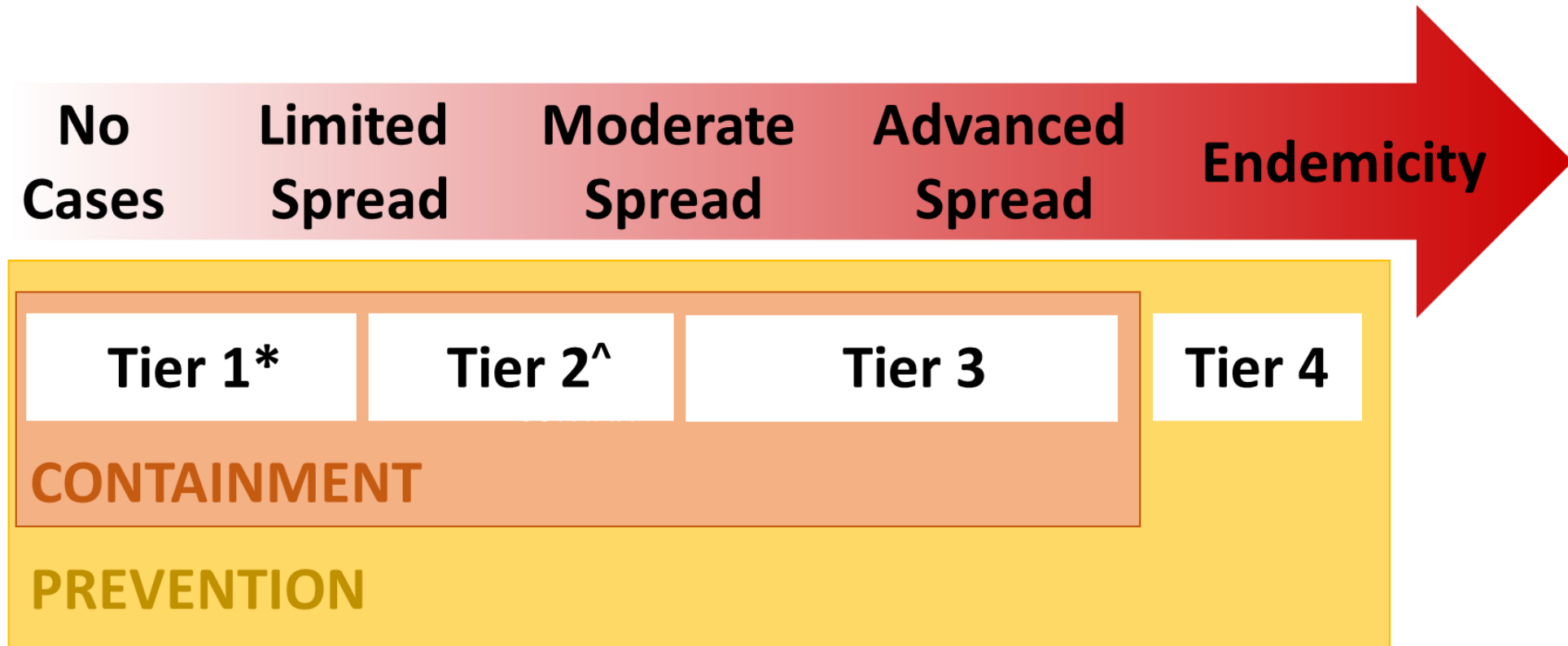
- Response to a **single case** of targeted resistance
- Goal to slow the spread of resistance
- 4-tiered approach based on organism/mechanism and local epidemiology

Tier	Description	Michigan Examples
1	Novel Resistance and/or resistance mechanisms never or rarely identified in the U.S.	Novel organism VRSA
2	Found in healthcare settings but not found regularly; No current treatment options exist and potential to spread more widely.	Any CPO with NDM, OXA-48, VIM, IMP CRPA or CRAB with KPC <i>Candida auris</i> Pan-Nonsusceptible (I or R to all drugs tested) organisms
3	MDROs targeted by region, but not considered endemic.	CP-CRE with KPC or CRAB OXA-23 (Regions 7/8)
4	Endemic in a region	CP-CRE with KPC or CRAB OXA-23 (Other Regions)



[CDC Containment Strategy Guidelines for Targeted MDROs](#)

Tier Definitions, Epidemic Stages, Response and Prevention



Organisms or resistance mechanisms that have

*Never (or very rarely) been identified **in the United States** and for which experience is extremely limited are Tier 1

^ Never (or very rarely) been identified **in a public health jurisdiction but are more common in other parts of the U.S.** are Tier 2.

2022 Update: Containment Response Elements

		Tier 1	Tier 2	Tier 3
Healthcare investigation	Review the patient's healthcare exposures prior to and after the positive culture	30 days	30 days	Current, sometimes prior admission
Contact investigation	Screen healthcare roommates			
	Screen additional healthcare contacts			
	Screen household contacts			
	Screen healthcare personnel			
If transmission identified	Repeat PPS at regular intervals if cases identified*			
	Evaluate potential for spread to linked facilities			
Clinical surveillance	Prospective laboratory surveillance			
	Retrospective laboratory surveillance			
Environmental cx	Environmental Sampling			
Ensure adherence to IPC	Infection control assessment w/ observations of practice			

	ALWAYS
	USUALLY
	SOMETIMES
	RARELY

*Periodic (e.g., every two weeks) response-driven PPS should be conducted until transmission is controlled, defined as two consecutive PPS with no new cases identified or, in facilities with high colonization pressure, substantially decreased transmission. If high levels of transmission persist across multiple point prevalence surveys in long term care settings, consider increasing the interval between surveys or temporarily pausing them while reassessing infection control and implementing interventions.

VRSA Investigation Steps

- 1) Develop a plan for VRSA colonized or infected patients
- 2) Identify and categorize contacts
- 3) Specimen collection and screening
- 4) Evaluate Efficacy of Infection Control Precautions

https://www.cdc.gov/hai/pdfs/VRSA-Investigation-Guide-05_12_2015.pdf



Investigation and Control of Vancomycin- Resistant *Staphylococcus aureus* (VRSA): 2015 Update

Division of Healthcare Quality Promotion
Centers for Disease Control and Prevention
Updated: April 2015



DEPARTMENT OF HEALTH AND HUMAN SERVICES
CENTERS FOR DISEASE CONTROL AND PREVENTION

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New ICAR Tool

What is an ICAR?

- Infection Control Assessment and Response (ICAR)
- Systematic assessment of a facility's IPC practices
 - Identifies gaps in practices
 - Guides quality improvement
- ICAR tool for general IPC across settings
 - Acute care, long-term care, and outpatient settings
 - Series of 10 modules that can be selected for use by ICAR facilitator
 - Modules are not setting specific

Containment Response

Case Study #1: CP-CRE

Case Study #1

BOL Lab Notification:
NDM+ *Klebsiella pneumoniae*
L hip wound
SNF A resident

SNF A Screened 2 of
23 recommended
contacts = negative
Declined ICAR

Feb 2021

Mar 2021

Apr 2021

May 2021

June 2021

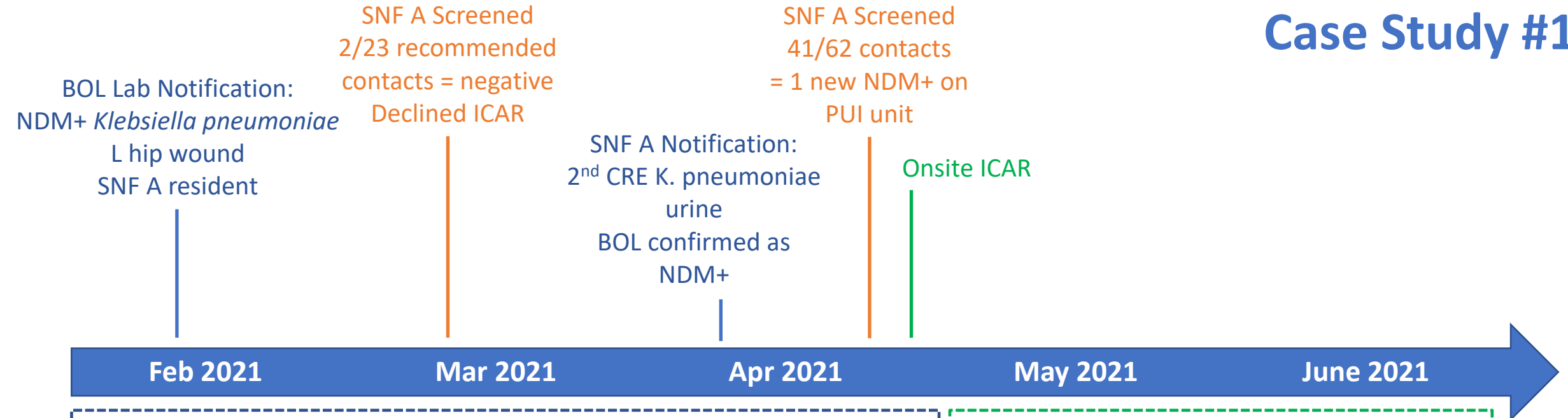
Case Investigation Found:

- Resided on COVID-19 PUI unit since Oct 2020
- Received in room wound care, PT/OT
- SNF A practicing extended use/reuse of PPE and experiencing staffing shortages

Provided IP Recommendations:

- Enhanced Barrier Precautions
- Transition away from extended use/reuse of PPE
- Ensure high adherence to IPC practices
- Conduct CP-CRE colonization screening for healthcare contacts on PUI unit
- Participate in an ICAR

Case Study #1



Case Investigation Found:

- Resided on LTC unit since Aug 2018, last hospitalization May 2019
- Colostomy that sometimes leaks, dementia
- SNF A still practicing extended use/reuse of PPE and experiencing staffing shortages

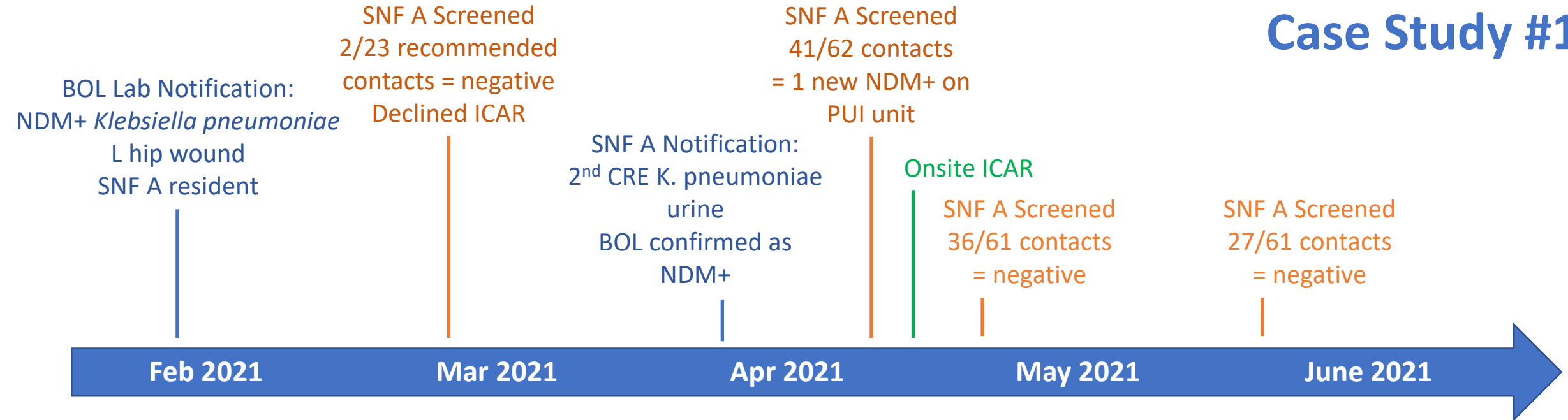
Provided IP Recommendations:

- Enhanced Barrier Precautions
- Transition away from extended use/reuse of PPE
- Ensure high adherence to IPC practices
- Conduct CP-CRE colonization screening for healthcare contacts on all units
- Participate in an ICAR

Onsite ICAR:

- Enhanced Barrier Precautions not fully implemented
- Still practicing extended/reuse PPE
- PPE supplies not stored near point-of-use
- ABHS not available in resident rooms, missed opportunities for hand hygiene observed
- Observed gaps in cleaning & disinfection practices

Case Study #1



Enhanced Surveillance:

- No additional cases detected from clinical cultures at SNF A
- Hospital A (shares patients)
 - Did not detect any additional cases
 - Provided IP recommendations

Case Study #1 Recap

Containment Response Activities

Element	Activity	Tier 2 Recommendation	Case Study #1
Healthcare investigation	Review the patient's healthcare exposures prior to and after the positive culture	30 days	Cases #1, 2, 3
Contact investigation	Screen healthcare roommates	✓	LTC
	Screen additional healthcare contacts	✓	PUI & LTC
	Screen household contacts	X	X
	Screen healthcare personnel	X	X
If transmission identified	Repeat PPS at regular intervals if cases identified*	✓	4 PPS
	Evaluate potential for spread to linked facilities	✓	Hospital A
Clinical surveillance	Prospective laboratory surveillance	✓	Monitored
	Retrospective laboratory	✓	Reviewed
Environmental cx	Environmental Sampling	X	X
Ensure adherence to IPC	Infection control assessment w/ observations of practice	✓	Onsite ICAR

Containment Response

Case Study #2: *C. auris*

Case Study #2

Clinical Lab
Notification:
Candida auris
urine
LTACH A patient

Screened 19
patients = 3 CA+

Mar 2022

Apr

May

June

July

Aug

Sept

Case Investigation Found:

- Admitted to LTACH A since Jan 2022; other recent HCF exposures at ACH, LTACH, vSNF since Jul 2021
- Chronic trach/vent, PEG, foley, midline IV, chronic wounds
- On contact precautions since admission

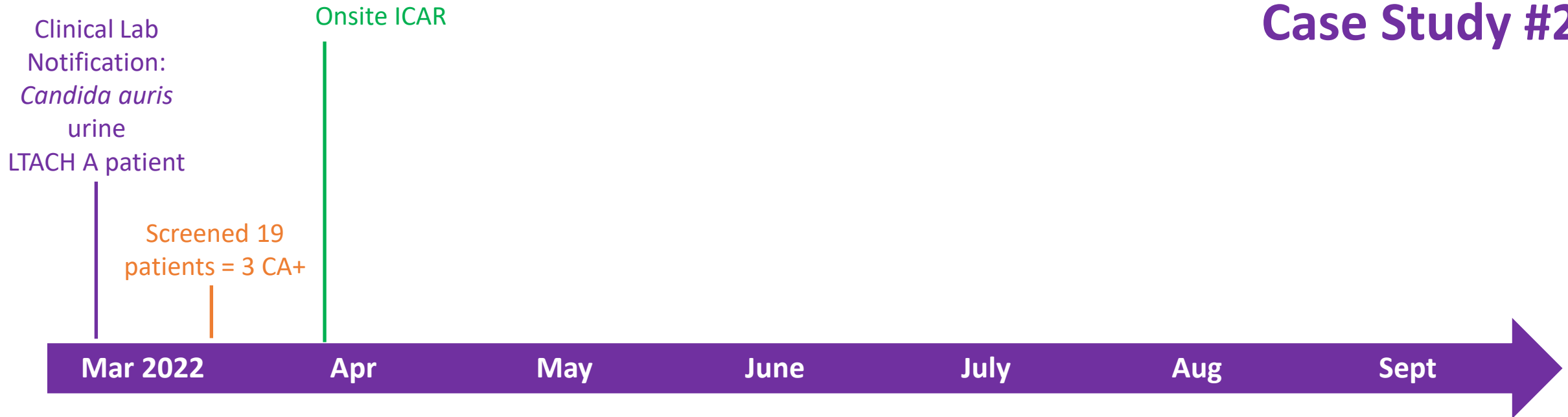
Provided IP Recommendations:

- Contact Precautions
- Ensure high adherence to IPC practices
- Use an EPA List P disinfectant
- Conduct *C. auris* colonization screening for all patients at facility
- Participate in an ICAR

Further IP Recommendations:

- Barrier Precautions for all patients
- Use an EPA List P disinfectant for whole facility
- Conduct *C. auris* & CPO colonization screening for all patients at facility every 2 weeks; admission & discharge screening
- Notify and screen discharges to HCF in past 30 days

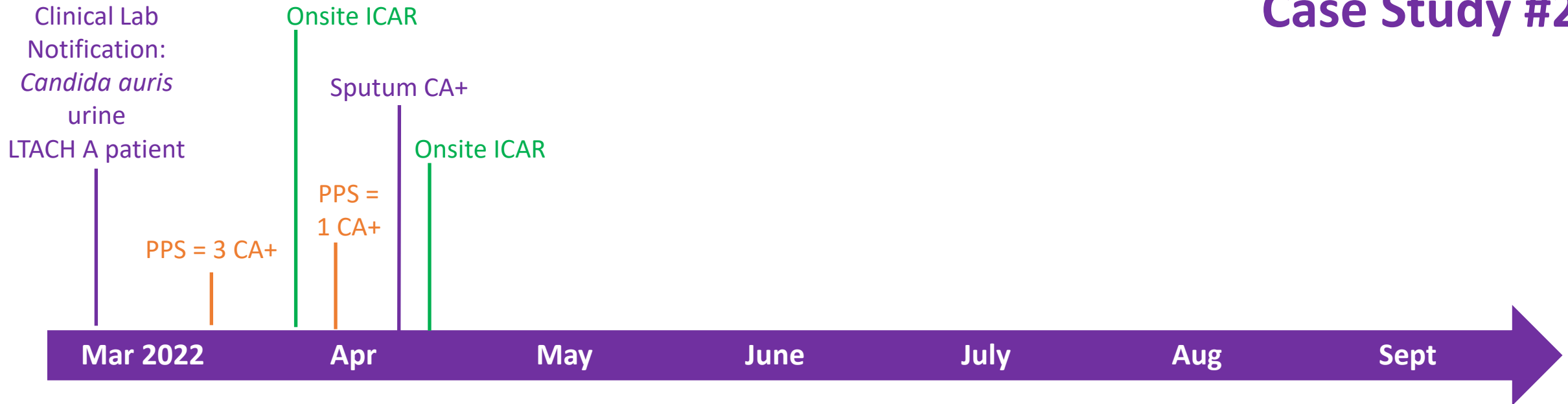
Case Study #2



Onsite ICAR:

- Reviewed outbreak action plan with leadership
- Review case healthcare records, exposure histories, and room/bed movement
- Discussed IPC policies/procedures
- Observations of IPC practices
- Reviewed disinfectant products available – currently only List K disinfectants
- Provided recommendations to strengthen auditing/feedback for hand hygiene, PPE, cleaning & disinfection

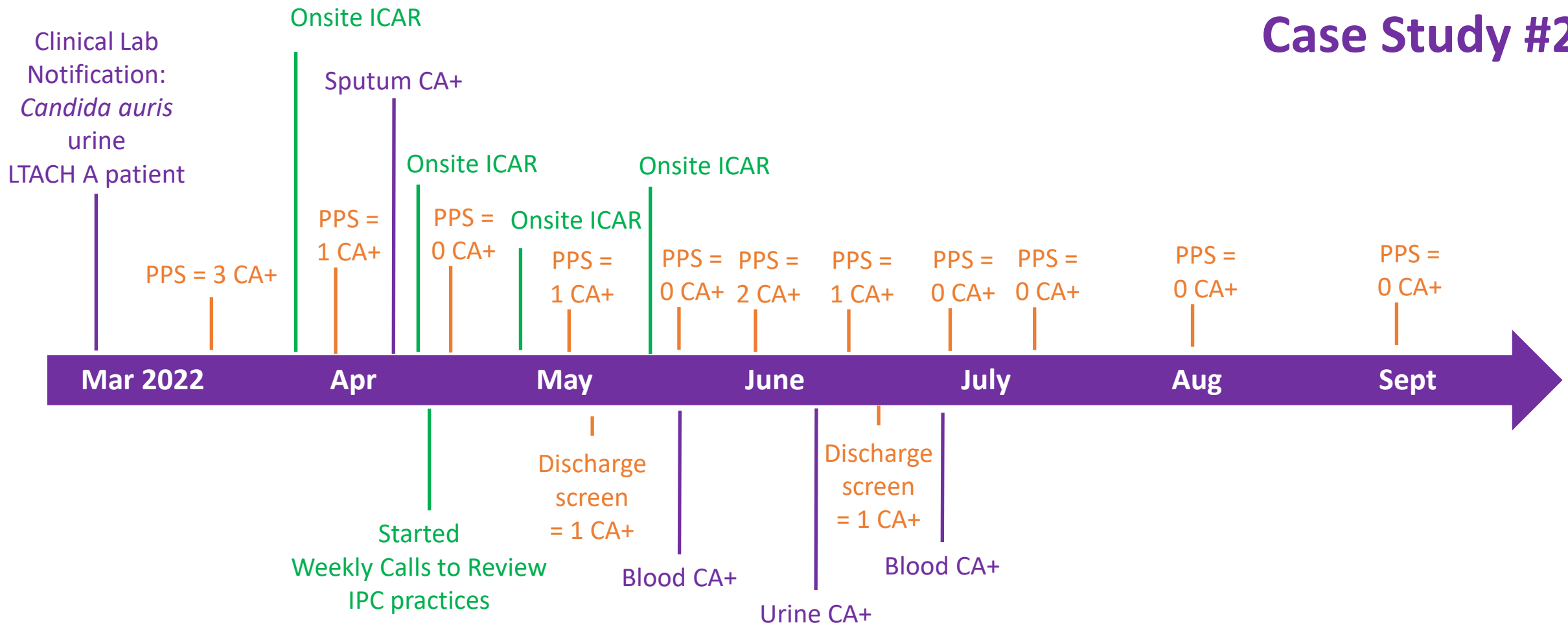
Case Study #2



Onsite ICAR:

- Reviewed outbreak action plans
- Observations of IPC practices
 - Missed opportunities for hand hygiene, disinfection of mobile workstations for clinical documentation and medication administration
- Reviewed disinfectant products available – mix of List P & List K disinfectants

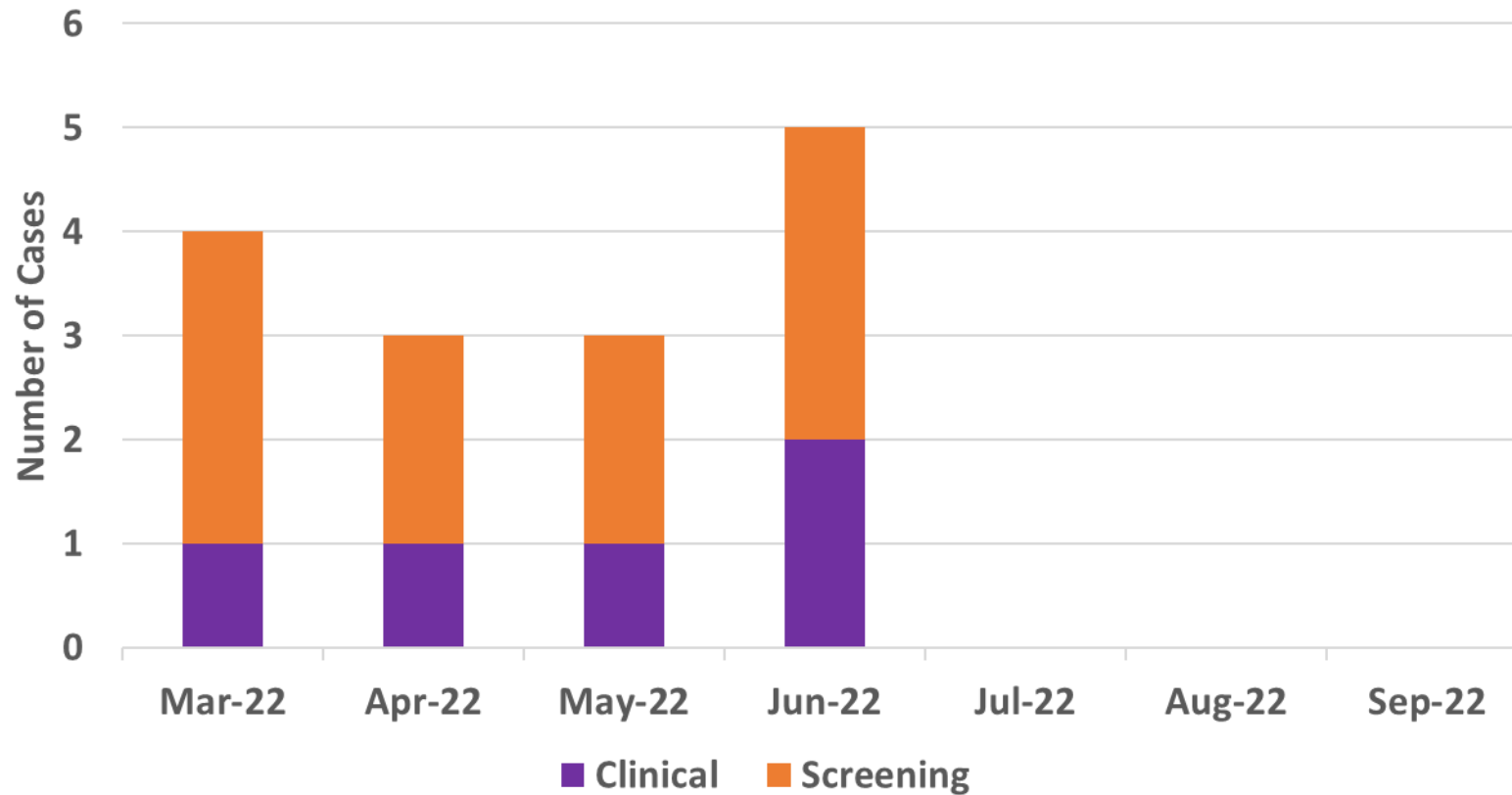
Case Study #2



Apr 2022 - Colonization screening conducted at epi-linked HCF related to index patient & recent discharges = no additional cases

Case Study #2 Recap – Epi Curve

Candida auris Cases at LTACH A



- 15 *C. auris* cases detected in 12 patients
 - 2 clinical cultures
 - 3 initially detected on colonization screening with subsequent clinical cultures
 - 10 colonization screening

Case Study #2 Recap

Containment Response Activities

Element	Activity	Tier 2 Recommendation	Case Study #2
Healthcare investigation	Review the patient's healthcare exposures prior to and after the positive culture	30 days	15 cases in 12 patients
Contact investigation	Screen healthcare roommates	✓	All roommates
	Screen additional healthcare contacts	✓	All patients
	Screen household contacts	X	X
	Screen healthcare personnel	X	X
If transmission identified	Repeat PPS at regular intervals if cases identified*	✓	11 PPS
	Evaluate potential for spread to linked facilities	✓	Epi-linked HCF and Discharges 30 days prior to index
Clinical surveillance	Prospective laboratory surveillance	✓	Monitored – 4 more clinical cases
	Retrospective laboratory	✓	Reviewed
Environmental cx	Environmental Sampling	X	X
Ensure adherence to IPC	Infection control assessment w/ observations of practice	✓	4 Onsite ICARs plus weekly calls

Questions?

Contact:
MDHHS SHARP Unit

